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# Indonesian red chilli (*Capsicum annuum* L.) capsaicin and its correlation with their responses to pathogenic *Fusarium oxysporum*

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### ABSTRACT

Red chili is a commercial crop for the food industry in Indonesia. There are some categories of red chili based on their pungency. The hot chili usually has more capsaicin than the sweet chili. Some cultivars may have more resistance to pathogen infection than the others. This research aimed to analyze the disease resistance of red chili cultivars from Indonesia against pathogenic *Fusariumoxysporum* and the correlation with capsaicin contents. Disease resistance was examined by determination of the Disease Severity Index (DSI) 15 dpi (days post inoculation). The correlation was analyzed by the regression coefficient. The result showed that the most resistance cultivar against *F. oxysporum* was Branang, while Lembang-1displayed the contrary. There was not a correlation of capsaicin content with the chili resistance to *F. oxysporum*.

Keywords: capsaicin, disease severity index, chili

#### I. INTRODUCTION

Chilli (*Capsicum*) is one of the family Solanaceae. It is from South America (7.500 BC) which have about 25 wild species as the progenitor (Perry et al., 2007). Now there are five species domesticated, includes *C. annuum*, *C. frutescens*, *C. chinense*, *C. baccatum*, and *C. pubescens* (Pickersgill, 1997). The domesticated species came to Asia by Portugal and Spain trading and were dispersed mainly to Philippine, India, China, Indonesia, Korea, and Japan (Perry et al., 2007).

Indonesia has domesticated many cultivars of *C. annuum* that known as red chili or big chili. Indonesian used the chili as a spice of their food. The food industry uses the red chili as a raw material for chili sauce and chili-powder products. The medical industry uses red chili as a capsaicin source for pain treatment. The *C. annuum* was categorized into two varieties, namely as *C. annuum* variety longum and *C. annuum* variety grossum. Indonesian mostly define *C. annuum* variety as longum. The *C. annuum* variety grossum was recognized as "paprika" and just found in the high and cold area (Djarwaningsih, 2005). The red chili was cultivated from the landrace to the mountain as an annual crop (Setiadi, 2011). There are big red chili and curly-red chili based on the difference in fruit surface. The big red chili has a smooth surface, while the curly-red chili has a wrinkled surface of the fruit. There were 86 cultivars of big-red chili and 87 cultivars of curly-red chili that registered in the Agriculture Ministry of Indonesia by 2011. This research aimed to analyze the disease resistance of red chili cultivars from Indonesia against pathogenic *Fusarium oxysporum* and the correlation with capsaicin contents.

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#### **II. MATERIAL AND METHODS**

#### **Fungal and Plant materials**

Pathogenic *Fusariumoxysporum* was isolated from wilting fusarium chili in Tawangmangu, Karanganyar Indonesia (Ferniah, *et al.*, 2014). The fungi were grown in Potato Dextrose Agar and Broth for the cultivation before inoculated to plants. This research used local Indonesian red-chili cultivars. Branang, Gantari, and Cipanas were open-pollinated cultivars produced by Indonesian Breeding Centers. Lembang-1 andKencana were open-pollinated cultivars produced by Indonesian Vegetable Research Centre.

#### Methods

Capsaicin content was analyzed for each cultivar of chili. The analysis was done by research service center (LPPT) of Gadjah Mada University, Yogyakarta Indonesia, using thin layer chromatography.

Seeds were spread in a tray and grown under plastic canopy, one tray to one cultivar. After 7 - 10 days the seedlings started to grow. Then the seedlings were planted into small polybag (3 x 5 cm) contains topsoil and maintained under plastic canopy. On 30 days after planting (dap), each cultivar was grown in 30 x 30 cm polybag contained topsoil and maintained carefully. The experiment was completely randomized design with ten replicates of each cultivar.

*Fusarium oxysporum* was grown in Potato Dextrose Broth (PDB) for four daysand incubated up to  $10^6$  conidia/mL. The conidia were inoculated on 30-day-old chili plants by the root dip method (Herman & Perl-Treves, 2007; Karimi *et al.*, 2010). Disease symptoms were observed every other day post-inoculation (dpi) for 15 dpi. Symptoms were recorded using the following system: Score 0 = no symptom, 1 = lower height compared to control, 2 = lower height and chlorosis, 3 = 10% chlorosis and/or 10% wilting, 4 = 11-25% wilting, 5 = 26-50% wilting, 6 = 51-100% wilting and dead. The disease severity index (DSI) was determined by the following equation (Wongpia & Lomthaisong, 2010):

 $DSI = \sum \frac{\text{(Disease severity scale x number of plants in each scale)}}{\text{(Highest numerical scale index x total number of plants)}} \times 100\%$ 

Based on their DSI, plants were categorized as highly resistant (HR) if  $0\% < DSI \le 2\%$ , resistant (R) if  $2\% < DSI \le 10\%$ , susceptible (S) if  $10\% < DSI \le 30\%$ , and highly susceptible (HS) if  $30\% < DSI \le 100\%$  (modified from Nsabiyera *et al.*, 2012).

Correlation of the capsaicin content and disease severity index was analyzed by correlation curve.

#### **III. RESULTS AND DISCUSSION**

Capsaicin content of Indonesian red chili showed the variable amount. It is accordance with Nwokem *et al.* (2010) that determine many variable capsaicin contents from Nigerian chili. Table 1 showed Indonesian red chili capsaicin content.

Table 1. Capsaicin content of Indonesian red chilli cultivars	
Cultivar	Capsaicin content (mg/100
	g)
Cipanas	0.923
Lembang-1	0.779
Branang	0.744
Gantari	0.712
Kencana	0.430

The table showed that Indonesian red chili has more capsaicin content (0.430 - 0.923 mg/g) than Nigerian chili (0.116 - 0.810 mg/g) based on Nwokem *et al.* (2010) research. Usually, the capsaicin content is correlated with the pungency. In Nigerian chili, the most pungent chili has the most capsaicin, and the less pungent chili has the less capsaicin. So, it is possible for Indonesian red chili to have more pungency than the Nigerian chili.



Figure 1. Disease Severity Index of Indonesian red chili cultivars inoculated with pathogenic *Fusarium oxysporum* at nine dpi (days post inoculation)

The resistance testing showed that Branang has the highest resistance to *F. oxysporum* infection. This is indicated by the smallest value of the Disease Severity Index (6.9%) compared to other chili cultivars. DSI values are shown in Figure 1. Based on the DSI values of each cultivar, Branang is categorized as resistant, Gantari and Cipanas are categorized as susceptible, while Lembang-1 and Kencana are classified as highly susceptible. The value of DSI and the resistance of chili plants is in accordance with the previous research indicating that Branang was a resistant cultivar and Lembang-1 was a cultivar of Highly Susceptible (Ferniah et al., 2014). Plant resistance is determined by the genetic differences of each cultivar and its adaptability to the environment.

The relationship between capsaicin content and DSI value can be seen from the regression correlation graph. Figure 2 shows that the regression coefficient (R2) is 0.128, which means that there is no good correlation between capsaicin content and DSI.



Figure 2. Capsaicin and DSI correlation of Indonesian red chilli

# **IV. CONCLUSION**

The most resistant cultivar against *F. oxysporum* was Branang, while Lembang-1 displayed the contrary. There was not a correlation of capsaicin content with the chili resistance to *F. oxysporum*.

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# REFERENCES

- Djarwaningsih, T. (2005).Capsicum spp.: Asal, PersebarandanNilaiEkonomi (Capsicum spp.: origin, distribution, and economic value. Biodiversitas6 : 292 296.
- Esbaugh, W.H. (1983). The Genus of Capsicum (Solanaceae) in the Bahamas.Proceeding of the second symposium on the botany of the Bahamas.
- IPGRI (1995).Descriptors for Capsicum (Capsicum spp.).International Plant Genetic Resources Institute, Rome, Italy; the Asian Vegetable Research and Development Center, Taipei, Taiwan, and the Centro Agronómico Tropical de Investigación y Enseñanza, Turrialba, Costa Rica.Pp. 110.
- Islam, M., Saha, S., Akand, M.D.H., and Rahim, Md.A. (2011). Effect of spacing on the growth and yield of sweet pepper (Capsicum annuum L.). Journal of Central European Agriculture. 12: 328 335.
- Kolekar, P. Kale, M., and Kulkarni-Kale, U. (2011). Molecular Evolution & Phylogeny: What, When, Why & How?, Computational Biology and Applied Bioinformatics, Prof. Heitor Lopes (Ed.), ISBN: 978- 953-307-629-4, InTech, Available from <u>http://www.intechopen.com/books/computational-biology-and-</u> appliedbioinformatics/molecular-evolution-phylogeny-what-when-why-how-
- Misra, S., Lal, R.K., Darokar, M.P., and Khanuja, S.P.S. (2011). Genetic Variability in Germplasm Accessions of Capsicum annuum L.American Journal of Plant Sciences. 2: 629 635.
- Nwokem, C.O., Agbaji, E.B., Kagbu, J.A., and Ekanem, E.J. 2010.Determination of Capsaicin Content and Pungency Level of Five Different Peppers Grown in Nigeria.New York Science Journal 3 (9).
- Pickersgill, B. (1997) Genetic resources and breeding of Capsicum spp. Euphytica 96:129–133.
- Perry, L., Dickau, R., Zarrillo, S., Holst, I., Pearsall, D.M., Piperno, D.R., Berman, M.J., Cooke, R.J., Rademaker, K., Ranere, A.J., Raymond, J.S., Sandweiss, D.H., Scaramelli, F., Tarble, K., and Zeidler, J.A. (2007). Starch Fossils and the Domestication and Dispersal of Chili Peppers (Capsicum spp. L.) in the Americas. Science 315: 986-988.
- Rego, E.R., Rego, M.M., Cruz, C.D., Finger, F.L., and Casali, V.W.D.(2011). Phenotypic diversity, correlation, and importance of variables for fruit quality and yield traits in Brazilian peppers (Capsicum baccatum). Genetics Resources Crops Evolution 58: 909 – 918.
- Rosmaina, Syafrudin, Hasrol, Yanti, F., Juliyanti, and Zulfahmi (2016).Estimation of variability, heritability and genetic advance among local chili pepper genotypes cultivated in peatlands.Bulgarian Journal of Agricultural Science. 22: 431–436.
- Setiadi (2011).BertanamCabai di Lahandan Pot (Chilli cultivation in land and a pot).PenebarSwadaya. Depok.40 pp.
- Walsh, B.M., and Hoot, S.B. (2001).Phylogenetic Relationships OfCapsicum (Solanaceae) Using DNA Sequences From Two Noncoding Regions: The Chloroplast Atpb-RbclSpacer Region And Nuclear waxy introns. International Journal of Plant Science 162: 1409–1418.
- Yatung, T., Dubey, R.K., Singh, V., and Upadhyay, G. (2014). Genetic diversity of chili (Capsicum annuum L.) genotypes of India based on morpho-chemical traits. Australian Journal of Crop Science 8: 97 102.
- Abdillah S, Wahida Ahmad R, Kamal Muzaki F, Mohd Noor N (2013b) Antimalarial activity of Neopetrosia exigua extract in mice. Journal of Pharmacy Research 6, 799-803.
- Alivy A, Nurhayati APD, Subagio I, et al. (2016) Composition of Sponges Associated with Mangrove in Tampora Beach, Situbondo, East Java Proceeding 3rd International Biology Conference (IBOC) 2016: Biodiversity and Biotechnology for Human Welfare 1, 11-16.
- Bavestrello G, Bonito M, Sarà M (1993) Influence of depth on the size of sponge spicules. Scientia Marina(Barcelona) 57, 415-420.
- Becking LE, Cleary DFR, de Voogd NJ (2013) Sponge species composition, abundance, and cover in marine lakes and coastal mangroves in Berau, Indonesia. Marine Ecology Progress Series 481, 105-120.

Bell JJ (2008) The functional roles of marine sponges. Estuarine, Coastal and Shelf Science 79, 341-353. Bell JJ, Barnes DKA (2000) The influences of bathymetry and flow regime upon the morphology of sublittoral

sponge communities. Journal of the Marine Biological Association of the United Kingdom 80, 707-718. Bell JJ, Smith D, Hannan D, et al. (2014) Resilience to disturbance despite limited dispersal and self-recruitment in tropical barrel sponges: implications for conservation and management. PLoS One 9, e91635.

Briggs JC (1999) Coincident biogeographic patterns: Indo-West Pacific ocean. Evolution 53, 326-335.

- Calcinai B, Bastari A, Makapedua DM, Cerrano C (2016) Mangrove sponges from Bangka Island (North Sulawesi, Indonesia) with the description of a new species. Journal of the Marine Biological Association of the United Kingdom 97, 1417-1422.
- Cleary DFR, de Voogd NJ (2007) Environmental associations of sponges in the spermonde archipelago, Indonesia. Journal of the Marine Biological Association of the United Kingdom 87, 1669-1676.
- de Goeij JM, van Oevelen D, Vermeij MJA, et al. (2013) Surviving in a marine desert: the sponge loop retains resources within coral reefs. Science 342, 108-110.
- de Voogd NJ, Cleary. DFR, Hoeksema. BW, Noor. A, van Soest RWM (2006) Sponge beta diversity in the spermonde archipelago, SW Sulawesi, Indonesia. Marine Ecology Progress Series 309, 131-142.
- Diaz MC, Rützler K (2009) Biodiversity and abundance of sponges in Caribbean mangrove: indicators of environmental quality. Smithsonian Contributions of Marine Science 38, 151-172.
- Dohrmann M, Wörheide G (2013) Novel scenarios of early animal evolution—is it time to rewrite textbooks? Integrative and Comparative Biology 53, 503-511.
- Faulkner J (2002) Marine natural products. Natural Product Report 19, 1-48.
- Faunce CH, Serafy JE (2006) Mangroves as fish habitat: 50 years of field studies. Marine Ecology Progress Series 318, 1-18.
- Frøhlich H, Barthel D (1997) Silica uptake of the marine sponge Halichondria panicea in Kiel Bight. Marine Biology 128, 115-125.
- Hoeksema B (2004) Biodiversity and the natural resource management of coral reefs in Southeast Asia. In: Challenging coasts: transdisciplinary excursions into integrated coastal zone development (ed. Visser L), pp. 49–71. Amsterdam University Press.
- Hooper JNA, van Soest RWM (2002) Systema Porifera: a guide to the classification of sponges Kluwer Academic/Plenum Publishers.
- Maldonado M, Aguilar R, Bannister RJ, et al. (2015) Sponge Grounds as Key Marine Habitats: A Synthetic Review of Types, Structure, Functional Roles, and Conservation Concerns. In: Marine Animal Forests: The Ecology of Benthic Biodiversity Hotspots (eds. Rossi S, Bramanti L, Gori A, Orejas Saco del Valle C), pp. 1-39. Springer International Publishing, Cham.
- Noor YR, Khazali M, I NN S (1999) Panduan pengenalan mangrove di Indonesia PKA/WI-IP (Wetlands International-Indonesia Programme).
- Pisani D, Pett W, Dohrmann M, et al. (2015) Genomic data do not support comb jellies as the sister group to all other animals. Proceedings of the National Academy of Sciences of the United States of America 112, 15402-15407.
- Proksch P, Edrada R, Ebel R (2002) Drugs from the seas current status and microbiological implications. Applied Microbiology and Biotechnology 59, 125-134.
- Redjeki S (2013) Komposisi dan Kelimpahan Ikan di Ekosistem Mangrove di Kedungmalang, Jepara (Fish Community Structure in Mangrove Ecosystem at Kedung Malang, Jepara Regency). ILMU KELAUTAN: Indonesian Journal of Marine Sciences 18, 54-60.
- Setiawan E, Nurhayati APD, Muzaki FK (2009) Sponge diversity at Pecaron Bay Situbondo based on macroscopic and microscopic observation. IPTEK The Journal for Technology and Science 20.
- Swierts T, Peijnenburg KTCA, de Leeuw C, et al. (2013) Lock, stock and two different barrels: comparing the genetic composition of morphotypes of the Indo-Pacific sponge Xestospongia testudinaria. PLoS One 8, e74396.
- van Soest RWM, Boury-Esnault N, Hooper JNA, et al. (2018) World Porifera database. http://www.marinespecies.org/porifera

- van Soest RWM, Boury-Esnault N, Vacelet J, et al. (2012) Global diversity of sponges (Porifera). PLoS One 7, e35105.
- Wörheide G, Erpenbeck D (2007) DNA taxonomy of sponges progress and perspectives. Journal of the Marine Biological Association of the United Kingdom 87, 1629-1633.

Wörheide G, Erpenbeck D, Menke C (2007) The sponge barcoding project – aiding in the identification and description of poriferan taxa. In: Museu Nacional Serie Livros (eds. Custódio MR, Hajdu E), pp. 123-128.

Wulff JL (2004) Sponges on mangrove roots, Twin Cays, Belize: early stages of community assembly National Museum of Natural History, Smithsonian Institution.