**The effectivity of parsley (*Petroselinum crispum*) extract on the growth inhibition of *Candida albicans***

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**ABSTRACT**

**Background**: Denture stomatitis is a type of *Candida-*associated infection that mainly affects the palatal mucosa. *Candida albicans* is one of the normal flora that is considered to be the primary etiologic agent in the pathogenesis of denture stomatitis. Denture decontamination is necessary to prevent denture stomatitis. One method of decontamination is by immersing removable dentures in an antifungal solution. Parsley (*Petroselinum crispum*) is a medicinal plant showing antifungal activity. **Objectives**: To determine the effectivity of immersion of acrylic resin Plate in parsley extract on inhibition of growth of *Candida albicans*. **Methods**: The effectivity of immersion of heat-cured acrylic resin plates in parsley extract on inhibition of *C. albicans* growth was tested using an experimental laboratory study with a post-test-only control group design. Thirty samples were divided into 5 groups as 0.01%, 0.02%, and 0.05% parsley ethanol extract, 0.1% sodium hypochlorite, and sterile aquadest. **Results**: 0.01% parsley ethanol extract is MIC with a mean of 7.4 CFU / ml, which inhibited the growth of *C. albicans* by 31.05%. The Kruskal Wallis test (p<0.001) results indicate that there is an effectivity of immersion of acrylic resin plate in parsley extract on inhibition of growth of *Candida albicans*. **Conclusions**: The immersion of acrylic resin plates in parsley ethanol extract with a concentration of 0.01%, 0.02%, and 0.04% has effectivity on the inhibition of growth of *C. albicans.*

**Key** words: Parsley extract, Candida albicans, removable denture cleaning solution, MIC.

**INTRODUCTION**

Edentulous is the condition of having no teeth that cause trouble with chewing and speech.1 Data of Riskesdas 2018 showed that in Central Java, 17.35% of edentulous cases happened because of tooth extraction or tooth loss. In Central Java, edentulous occur for 21.93 percent of the population aged 35 to over 65.2 The use of removable dentures is indicated for patients with no teeth.3 The function of removable dentures is to replace the lost natural teeth, which the patients can wear and remove themselves.4

The material commonly used as a removable tooth base is heat-cured polymethyl methacrylate (PMMA) resin.5-6 The use of PMMA as a removable tooth base is advantageous because it is easy to manufacture, relatively inexpensive, stable in the oral environment, has color stability, physical properties, and pleasing aesthetics.6-7 This type of resin will be formed if the polymerization process is carried out by heating so that a mixture of methyl methacrylate monomer and methacrylate polymer occurs. This process always leaves as much as 0.2% to 0.5% methyl methacrylate because the monomer will not completely react after polymerization is complete.5-6 Residual monomers can cause irritation, inflammation, allergic responses to the oral mucosa, angular cheilitis, and denture stomatitis.5

Denture stomatitis is a type of *Candida-*associated infection that mainly affects the palatal mucosa. This disease is characterized by mucosal inflammation, burning sensation, bleeding, and pain, especially under the upper denture, and changes in the sense of taste.8–10 The risk factor increased in patients wearing partial dentures to 36.7% and increased to 65% in patients wearing complete dentures.9 *Candida albicans* is one of the normal flora that is considered to be the primary etiologic agent in the pathogenesis of denture stomatitis.8

Candidiasis can occur if the hygiene of removable dentures is poor. Therefore, decontamination of removable dentures is essential for the prevention of denture stomatitis.11–13 Decontamination of dentures can be done by mechanical methods, chemical methods, and a combination of the two methods. The release of chemical solutions can be an option as a chemical method.3 Sodium hypochlorite (0.10% and 0.22%) is one of the most used chemical solutions as denture cleaners because it is effective in reducing *Candida albicans* colonization, is non-abrasive, and can be used directly. However, this solution can cause discoloration on the plate and is corrosive.13 Another chemical solution that is often used is alkaline peroxide. Soaking the denture in alkaline peroxide triggers a diffusion process that will change the acrylic resin's physical properties, including its hardness.14 Natural antifungal solutions can be an alternative to removable denture cleaning solutions because of the side effect of chemical-based antifungal solutions as mentioned above.15

Parsley (*Petroselinum crispum*) is a medicinal and aromatic plant from the Mediterranean that has health benefits.16–18 This plant contains vitamins, flavonoids, essential components, minerals, phenolic compounds, and other bioactive compounds. Parsley leaves can reduce the risk of gastrointestinal disorders, diabetes, and cardiovascular disease due to their antioxidant, cytoprotective, and antidiabetic activities.18,19 Other studies have shown that parsley exhibits antibacterial, antifungal, anti-inflammatory, and anticancer activities.19

One of the compounds that show antifungal activity and is contained in parsley extract is a flavonoid.20,21 Flavonoid can break down protein bonds into their primary structure on the cell membrane of *C. albicans* and cause lysis of the fungal cell membrane, allowing the flavonoid can penetrate the nucleus cell. The presence of flavonoid in the cell nucleus can stop the growth of *C. albicans*.22 Some researchers are interested to develop novel medication because it contains a lot of flavonoids.21

From the explanation above, it is necessary to investigate further regarding the effectivity of immersing acrylic resin plates in parsley (*P. crispum*) extract on the growth potential of *C. albicans* as a basis for the development of a removable denture cleaning solution with natural base ingredients.

**MATERIALS AND METHODS**

This study was conducted at Diponegoro University’s Integrated Laboratory and Microbiology Laboratory from March to April 2021 using an experimental laboratory design with a post-test only control group. The material used in this study was parsley (*Petroselinum crispum*) ethanol extract with a concentration of 0.01%, 0.02%, and 0.04%, 0.01% sodium hypochlorite sodium as positive control and sterile aquadest as negative control groups. *Candida albicans* were obtained from a culture at the Microbiology Laboratory of Diponegoro University.

*Candida albicans* suspension was prepared by culturing the colonies in 10 ml of BHI media and incubated for 8 hours at 370C. Then the suspension of *C. albicans* is diluted by adding sterile distillated water to certain turbidity to the McFarland standard 3 x 108.

Parsley (*Petroselinum crispum*) taken from Nandisari Plantation was extracted with 96% ethanol solvent (EMSURE®; Merck Millipore, Germany) using the maceration method and diluted with sterile distilled water to obtain concentrations of 0.01%, 0.02%, and 0.04%. Sterile acrylic resin plates were immersed in artificial saliva AFNOR NF S91-141 for 1 hour and rinsed using Phosphate Buffered Saline (Nitra Kimia, Indonesia) 2 times. Then the acrylic plates were immersed in *Candida albicans* suspension with a turbidity of 0.5 McFarland and incubated for 24 hours at 37°C. A total of 30 acrylic resin plates were removed and divided into five treatment groups, namely immersion in 0.01%, 0.02%, and 0.04% parsley ethanol extract and the control group, 0.1% sodium hypochlorite, and aquadest solution, respectively 10 ml. The plates were immersed for 8 hours, allowing the serial dilution was implemented in each test tube to 10-3. The serial dilution of 0.01 ml was taken and tested on SDA media (Merck Millipore, Germany) and then incubated for 48 hours at 37°C. The number of colony forming units (CFU) per mL was obtained after calculating the colonies on SDA media, allowing the minimum inhibitory concentration (MIC) obtained according to the formula.

The data obtained were then analyzed by the Saphiro-Wilk test, Kruskal Wallis test followed by the Mann-Whitney test.



**Figure 1.** Parsley (*Petroselinum crispum*) taken from Nandisari Plantation

**RESULTS**

The plant harvested from Nandisari Plantation identified as *Petroselinum crispum (Mill.) Fuss* by the expert*.* The research continued with the phytochemical screening of parsley ethanol extract. As shown in table 1, the results of the Lieberman test, tannins, and flavonoids respectively showed red, green, and yellow colors after mixing the reagents for each test.

**Table 1.** Phytochemical screening results of parsley ethanol extract

|  |  |
| --- | --- |
| Test | Result |
| *Lieberman* | + |
| Tannins | + |
| Flavonoids | + |

The average and standard deviation of 6 groups of *Candida albicans* numbers were calculated after soaking acrylic resin plates in 0.01 percent, 0.02 percent, and 0.04 percent parsley extract, sterile aquadest, and 0.1 percent sodium hypochlorite, as shown in Table 2.

**Table 2.** The average and standard deviation of fungal number

|  |  |  |
| --- | --- | --- |
| Group | Fungal number average | Standard deviation |
|
| 0.01% Parsley Extract | 7.40 | 0.31 |
| 0.02% Parsley Extract | 3.75 | 0.24 |
| 0.04% Parsley Extract | 1.61 | 0.07 |
| Sterile Aquadest | 10.73 | 0.59 |
| 0.01% Sodium Hypochlorite | 0 | 0.00 |



**Figure 2.** *Candida albicans* colony count after 48 hours of incubation

In parsley extract groups, it can be seen that 0.04% parsley extract has the least number, namely 1.61 CFU/ml, while 0.01% parsley extract has the highest average number of fungi, namely 7.40 CFU/ml

The average number of fungi for each parsley extract concentration group (*Petroselinum crispum*) obtained was then used in the MIC calculation, which was carried out using a comparison with the sterile aquadest group. The results of the MIC calculation are listed in table 3 using the formula.

**Tabel 3.** MIC calculation results

|  |  |  |
| --- | --- | --- |
| No. | Treatment group | MIC |
| 1. | 0.01% Parsley Extract | 31.05% |
| 2. | 0.02% Parsley Extract | 65.06% |
| 3. | 0.04% Parsley Extract | 84.93% |

The results of the Shapiro-Wilk test on fungal numbers data from parsley extract groups of 0.01%, 0.02%, and 0.04% and sterile aquadest showed that the data were normally distributed (p>0.05). In comparison, the fungal number of 0.1% sodium hypochlorite group showed that the data were not normally distributed (p<0.05). The study continued with the Kruskal Wallis test due to one of the treatment groups showed that the data were not normally distributed.

The Kruskal Wallis test analysis results in table 4 show that the five treatment groups in this study got p-values < 0.001 (p < 0.05). Thus, it can be said that there is a significant difference in the fungal number.

**Tabel 4.** Kruskall Wallis test result

|  |  |  |  |
| --- | --- | --- | --- |
|  | Treatment group | Mean Rank | *p-*value |
| *Candida albicans* number | 0.01% Parsley Extract | 21.50 | <0.001 |
| 0.02% Parsley Extract | 15.50 |
| 0.04% Parsley Extract | 9.50 |
| Sterile Aquadest | 27.50 |
| 0.1% Sodium Hypochlorite | 3.50 |

The analysis continued using the Mann-Whitney test, which give an insight that all groups had differences in each group with a significance as shown in table 5.

**Tabel 5.** Mann Whitney test result

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  | 0.01% Parsley Extract | 0.02% Parsley Extract | 0.04% Parsley Extract | Sterile Aquadest | 0.1% Sodium Hypochlorite |
| 0.01% Parsley Extract |  | 0.004 | 0.004 | 0.004 | 0.002 |
| 0.02% Parsley Extract | 0.004 |  | 0.002 | 0.004 | 0.002 |
| 0.04% Parsley Extract | 0.004 | 0.002 |  | 0.004 | 0.002 |
| Sterile Aquadest | 0.004 | 0.004 | 0.004 |  | 0.002 |
| 0.1% Sodium Hypochlorite | 0.002 | 0.002 | 0.002 | 0.002 |  |

**DISCUSSION**

This study obtained the average number of *Candida albicans* fungi in the immersion of heat-cured acrylic resin plates in parsley extract concentrations of 0.01%, 0.02%, and 0.04%, 0.1% sodium hypochlorite, and sterile distilled water during 8 hours. The MIC was calculated using a comparison with the sterile aquadest group.24 Parsley extract at concentrations of 0.01%, 0.02% and 0.04% respectively showed MIC of 31.05%, 65.06% and 84.93%.

According to the data analysis the data showed a significant difference between positive control values, negative control, 0.01%, 0.02%, and 0.04% parsley extract (*P. crispum*). These results are in accordance with the research hypothesis that there is an effectivity of immersing acrylic resin plates in parsley extract on the growth inhibition of *C. albicans*.

In the 0.1% sodium hypochlorite group, no *C. albicans* colonies were found on SDA media because this solution effectively killed *C. albicans*. At the same time, sterile aquadest could not inhibit the growth of *C. albicans* to obtain the highest average number of fungi in the group among all groups*.*13The 0.01% parsley extract group showed the highest average fungal number of 7.4 CFU/ml, while the 0.04% parsley extract group showed the lowest average fungal number, namely 1.61 CFU/ml. The less the number of molds formed, the higher the antifungal activity of a substance.25 Immersion of acrylic resin plates in the 0.01%, 0.02%, and 0.04% ethanol extract of parsley (*P. crispum*) showed that the average number of fungi *C. albicans* were lower in numbers compared to the sterile aquadest group. This was caused by antifungal activity in parsley (*Petroselinum crispum*) extract.20,21,26 In the parsley (*P. crispum*) ethanol extract group, the number of fungi decreased with each increase in the concentration of parsley (*P. crispum*) ethanol extract. The greater the concentration of parsley extract, the greater the ability to inhibit fungal growth.26 This can occur following the basic chemical theory, which states that the greater the concentration of an antimicrobial substance, the faster microbial cells will be stunted and killed.27

From the MIC calculation, it can be seen that the three concentrations of parsley ethanol extract (0.01%, 0.02%, and 0.04%) inhibit *C. albicans* growth on acrylic resin plates and are fungistatic. According to Washington's opinion, if the MIC is less than 99.9%, then the solution is fungistatic.24 Although the three concentrations of parsley extract (*P. crispum*) can inhibit *C. albicans*, not all concentrations are effective as inhibitors of *C. albicans* on an acrylic plate. The ability of an antifungal agent can be said to be effective if it has a MIC of 80% or more when compared to the control.24 It can be concluded that 0.04% parsley ethanol extract effectively inhibits the growth of *C. albicans* on acrylic resin plates because it has a MIC of more than 80%, which is equal to 84.93%. However, in the concentration group of 0.01% and 0.02%, it could not effectively inhibit the growth of *C. albicans* on acrylic resin plates because the MICs were only 31.05% 65.05%, respectively.

The antifungal activity of parsley (*P. crispum*) extract is due to biochemical compounds in this plant.28 Liberal et al. (2020) stated that parsley contains bioactive compounds such as flavonoids which have been shown to have the ability to inhibit fungal growth.19,29 According to Alcamo, flavonoids are a class of phenolic compounds with fungistatic properties which support the previous statement.24 Phytochemical screening carried out in this study also showed the presence of flavonoids in the ethanol extract of parsley (*P. crispum*) due to a yellow color change when the extract was mixed with magnesium powder and amyl alcohol.30

According to Aboody and Mickymaray (2020), flavonoid compounds can inhibit fungal growth by various mechanisms, namely by damaging the fungal plasma membrane, damaging mitochondria, and inhibiting cell wall formation, cell division, RNA, and protein synthesis in fungi.31 Flavonoid can also break down protein bonds into their primary structure on the cell membrane of *C. albicans* and cause lysis of the fungal cell membrane, flavonoids can then enter the cell nucleus. The presence of flavonoid in the cell nucleus can stop the growth of *C. albicans*.22 Other studies have shown that flavonoid compounds in parsley such as apigenin can cause *Candida albicans* to experience cell shrinkage.32,33

There are other compounds that can inhibit the growth of fungi in parsley (*P. crispum*) extract, namely triterpenoid compounds.34 Phytochemical screening carried out in this study showed the presence of triterpenoids in the ethanol extract of parsley (*P. crispum*) due to a red color change when mixing the extract with acetic anhydride and concentrated H2SO4.30 This compound is a bioactive compound that has an antifungal function. Triterpenoids have a mechanism of decreasing the permeability of cell membranes of microorganisms and can be associated with protein and lipid molecules so that they can have an impact on the physiological functions of cell membrane proteins and enzyme proteins from fungi.35

The number of *Candida albicans* on SDA media after immersing heat-cured acrylic resin plates in parsley ethanol extract at a concentration of 0.04% showed the lowest number of fungi compared to concentrations of 0.01% and 0.02%, on the other hand the growth inhibition of *C. albicans* in 0.1% sodium hypochlorite showed 1.17 times more effective results than the 0.04% parsley (*P. crispum*) ethanol extract. This is a disadvantage of parsley ethanol extract (*P. crispum*) as a growth inhibitor of *C. albicans* on acrylic resin plates. Hence, further research is needed to increase the effectiveness of parsley (*P. crispum*) ethanol extract in inhibiting the growth of *C. albicans* on acrylic resin plates which can be done by increasing the concentration of the ethanolic extract of parsley (*P. crispum*).

**CONCLUSION**

Research on the effectivity of immersing acrylic resin plates in parsley (Pe*troselinum crispum*) extract on the growth inhibition of *Candida albicans* concluded that there was an effectivity of immersing acrylic resin plates in parsley (*P. crispum*) extract on the growth inhibition of *C. albicans*. In addition, in this study, 0.04% parsley ethanol extract was the best concentration in inhibiting the growth of *C. albicans* on acrylic resin plates.

This research is an initial study, and more research is needed to establish the concentration of parsley ethanol extract that is most effective in inhibiting the growth of *C. albicans* on acrylic resin plates at concentrations larger than 0.04 percent. Furthermore, it is also necessary to do qualitative phytochemical screening of parsley extract and redetermination of *C. albicans* on the culture results.

**ACKNOWLEDGMENT**

The Authors would like to thank Mr. Bambang Ismawan for his technical assistance during this study.

**REFERENCES**

1. Lee, D. J. & Saponaro, P. C. (2019). Management of Edentulous Patients. *Dent. Clin. North Am.* **63**, 249–261.

2. Riset Kesehatan Dasar. *Laporan Provinsi Jawa Tengah Riskesdas 2018*. (2018). *Kementerian Kesehatan RI*.

3. Ribeiro Rocha, G. dos S., Neves Duarte, T., de Oliveira Corrêa, G., Nampo, F. K. & de Paula Ramos, S. (2020). Chemical cleaning methods for prostheses colonized by Candida spp.: A systematic review. *J. Prosthet. Dent.* 1–6. doi:10.1016/j.prosdent.2019.10.004.

4. Driscoll, C. F., Freilich, M. A., Guckes, A. D., Knoernschild, K. L. & Mcgarry, T. J. (2017). The Glossary of Prosthodontic Terms: Ninth Edition. *J. Prosthet. Dent.* **117**, e1–e105.

5. Nejatian, T., Pezeshki, S. & Yaqin Syed, A. U. (2019). *Acrylic denture base materials*. *Advanced Dental Biomaterials* (Elsevier Ltd). doi:10.1016/b978-0-08-102476-8.00005-0.

6. Diansari, V., Fitriyani, S. & Haridhi, F, M. (2016).Studi Pelepasan Monomer Sisa Dari Resin Akrilik Heat Cured Setelah Perendaman Dalam Akuades. *Cakradonya Dent J* **8**, 1–76.

7. Alhareb, A. O., Akil, H. M. & Ahmad, Z. A. (2015). Mechanical Properties of PMMA Denture Base Reinforced by Nitrile Rubber Particles with Al2O3/YSZ Fillers. *Procedia Manuf.* **2**, 301–306.

8. Calderone, R. A. & Clancy, C. J. *Candida and Candidiasis*. (ASM Press).

9. Zainal, M., Mohamad Zain, N., Mohd Amin, I. & Ahmad, V. N. (2020). The antimicrobial and antibiofilm properties of allicin against Candida albicans and Staphylococcus aureus – A therapeutic potential for denture stomatitis. *Saudi Dent. J.* doi:10.1016/j.sdentj.2020.01.008.

10. Glick, M. & Feagans, W. *Burket’s Oral Medicine*. vol. 3. (2016). (People’s Medical Publishing House).

11. Gondim, B. L. C. *et al.* (2018). Effect of chitosan nanoparticles on the inhibition of Candida spp. biofilm on denture base surface. *Arch. Oral Biol.* **94**, 99–107.

12. An, S., Evans, J. L., Hamlet, S. & Love, R. M. Incorporation of antimicrobial agents in denture base resin : A systematic review. *J. Prosthet. Dent.* 1–8 doi:10.1016/j.prosdent.2020.03.033.

13. Arruda, C. N. F. de *et al.* (2017). Effect of sodium hypochlorite and Ricinus communis solutions on control of denture biofilm: A randomized crossover clinical trial. *J. Prosthet. Dent.* **117**, 729–734

14. Puspitasari, D., Saputera, D. & Anisyah, R. N. (2016). Perbandingan Kekerasan Resin Akrilik Tipe Heat Cured Pada Perendaman Larutan Desinfektan Alkalin Peroksida Dengan Ekstrak Seledri (Apium Graveolens L.) 75%). *ODONTO Dent. J.* **3**, 34

15. Deyab, M. H., Awady, B. E. & Bakir, N. G. (2018). Is immersion in mint oil or apple vinegar solution a valid antifungal approach for acrylic soft liners? *Futur. Dent. J.* **4**, 302–307

16. Derouich, M. *et al.* (2020). Assessment of total polyphenols, flavonoids and anti-inflammatory potential of three Apiaceae species grown in the Southeast of Morocco. *Sci. African* **9**, e00507.

17. Sbai, H., Saad, I., Ghezal, N., Greca, M. Della & Haouala, R. (2016).Bioactive compounds isolated from Petroselinum crispum L. leaves using bioguided fractionation. *Ind. Crops Prod.* **89**, 207–214.

18. Quan, Z. *et al.* (2020). Development and validation of a probe-based qPCR method to prevent parsley leaf material misidentification. *Fitoterapia* **146**, 104666.

19. Saleh, A. M., Selim, S., Jaouni, S. Al & AbdElgawad, H. (2018). CO 2 enrichment can enhance the nutritional and health benefits of parsley (Petroselinum crispum L.) and dill (Anethum graveolens L.). *Food Chem.* **269**, 519–526.

20. Jin, Y. S. (2019).Recent advances in natural antifungal flavonoids and their derivatives. *Bioorganic Med. Chem. Lett.* **29**.

21. Slimestad, R., Fossen, T. & Brede, C. (2020). Flavonoids and other phenolics in herbs commonly used in Norwegian commercial kitchens. *Food Chem.* **309**, 125678.

22. Supriyanto, Kuswiyanto & Nurhayati, E. (2017). Efektivitas Air Perasan Daun Lidah Buaya (Aloe Vera) Terhadap Pertumbuhan Jamur Trichophyton Rubrum Dengan Metode Dillu-tion Test. *Jamur Lab. Katulistiwa* **059**, 8–12.

23. Calderone, R. (2016). *Candida Species Methods and Protocols*. (Springer).

24. Rahman, E. (2021). Efektivitas Ekstrak Daun Dewa (Gynura pseudochina (Lour.) DC) terhadap Pertumbuhan Candida albicans pada Plat Dasar Gigi Tiruan Resin Akrilik. *Fak. Kedokt. Gigi UNISSULA* 1–13.

25. Abdu, K. & Hauwa, D. (2019). In vitro evaluation of the antioxidant and antimicrobial activity of leaf extracts of *Petroselinum cripsum* Parsley). *ChemSearch J.* **10**, 25–29.

26. Pineda, R., Vizcaíno, S., García, C. M., Gil, J. H. & Durango, D. (2018). Antifungal activity of extracts, essential oil and constituents from Petroselinum crispum against Colletotrichum acutatum. *Rev. Fac. Nac. Agron. Medellin* **71**, 8563–8572.

27. Sahrul, G. (2020). Uji Daya Hambat Ekstrak Minyak Atsiri Kayu Manis (Cinnamomun burmanii) terhadap Pertumbuhan Jamur Malassezia furfur. *JUSINDO* **2507**, 1–9 (2020).

28. Agyare, C., Appiah, T., Boakye, Y. D. & Apenteng, J. A. (2017). *Petroselinum crispum: A Review*. *Medicinal Spices and Vegetables from Africa: Therapeutic Potential Against Metabolic, Inflammatory, Infectious and Systemic Diseases* (Elsevier Inc.). doi:10.1016/B978-0-12-809286-6.00025-X.

29. Liberal, Â. *et al.* (2020). Bioactive Properties and Phenolic Compound Profiles of Turnip-Rooted, Plain-Leafed and Curly-Leafed Parsley Cultivars. *Molecules* **25**.

30. Purwati, S., Lumora, S. V. T. & Samsurianto. (2017). Skrining Fitokimia Daun Saliara (Lantana camara L) Sebagai Pestisida Nabati Penekan Hama dan Insidensi Penyakit Pada Tanaman Holtikultura di Kalimantan Timur. *Pros. Semin. Nas. Kim. 2017* 153–158.

31. Aboody, M. S. Al & Mickymaray, S. (2020). Anti-fungal efficacy and mechanisms of flavonoids. *Antibiotics* **9**.

32. Lee, H., Woo, E. R. & Lee, D. G. (2018). Apigenin induces cell shrinkage in Candida albicans by membrane perturbation. *FEMS Yeast Res.* **18**.

33. Aishwaya, J. (2018). Nutraceuticals Potential of Petroselinum Crispum: A Review. *J. Complement. Med. Altern. Healthc.* **7**, 1–6.

34. Daradkeh, G., Corporation, H. M. & Musthafa, M. E. (2018). Book chapter - parsley.

35. Komala, O., Yulianita & Fuji, R. S. (2019). Activity Of Antifungal Ethanol Extract 50% And Ethanol 96% Leaf Henna Nail (Lawsonia inermis L) Against Trichophyton mentagrophytes. *Ekol. J. Ilm. Ilmu Dasar dan Lingkung. Hidup* **19**, 12–19